

# Lirilumab Enhances Anti-Tumor Efficacy of Elotuzumab



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## Abstract

Tumor cells that express reduced levels of Major Histocompatibility Complex (MHC) class I molecules may be recognized and killed by Natural Killer cells (NK cells), through a process known as “missing self” recognition.

In humans, this is controlled by inhibitory receptors such as Killer Immunoglobulin-like Receptors (KIR) that recognize Human Leukocyte Antigen (HLA)-A, -B or -C. Engagement of KIR by HLA molecules results in inhibitory signaling that reduces NK cell-mediated natural killing and antibody dependent cellular cytotoxicity (ADCC). Hence, antibodies that block interactions between inhibitory KIR and their HLA ligands are being evaluated as an anti-cancer therapeutic strategy.

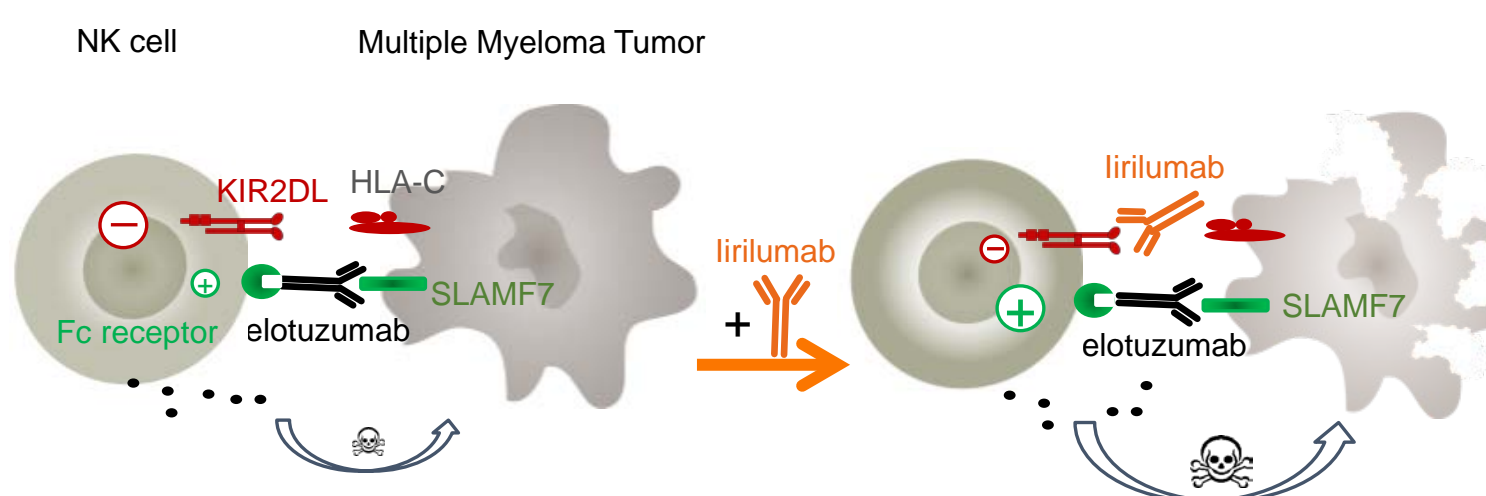
Lirilumab (BMS-986015 / IPH2102) is a fully human IgG<sub>4</sub> that blocks binding of inhibitory KIR to HLA-C, that is being developed for treating hematologic malignancies and solid tumors.

Elotuzumab (BMS901608 / HuLuc63) is a humanized IgG<sub>1</sub> anti-SLAMF7 (signaling lymphocyte activation molecule family member 7, CS-1) being developed for the treatment of Multiple Myeloma (MM). SLAMF7 is a cell surface glycoprotein highly expressed in myelomatous cells and only at low levels on normal cells.

NK cell-mediated ADCC is one of the main mechanisms of action of elotuzumab, but ADCC is negatively regulated by KIR checkpoint receptors. Thus a combination of lirilumab and elotuzumab has strong scientific rationale.

The aim of the present study was to assess whether lirilumab would enhance elotuzumab anti-MM activity *in vitro* with human peripheral blood NK cells and MM cell lines, and *in vivo* in a newly developed xenogenic mouse model.

## Background

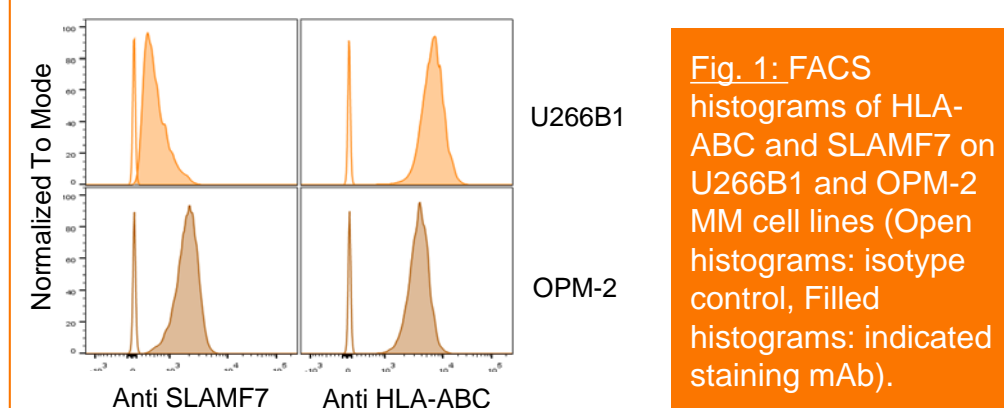


- ✓ NK cell inhibition by HLA-A/B/C
- ✓ elotuzumab-mediated ADCC negatively regulated by KIR checkpoint receptors
- ✓ Activation through KIR-HLA-blockade
- ✓ ADCC enhancement

## Materials & Methods

- Two MM cell lines (OPM-2 and U266B1) were identified that express both MHC Class I and SLAMF7.

**Figure 1: Expression of HLA-ABC and SLAMF7 on multiple myeloma cell lines U266B1 and OPM-2**



**Fig. 1:** FACS histograms of HLA-ABC and SLAMF7 on U266B1 and OPM-2 MM cell lines (Open histograms: isotype control, Filled histograms: indicated staining mAb).

- Functional *in vitro* assays were performed with peripheral blood mononuclear cells (PBMC) from healthy volunteers as effector cells (E/T ratio = 2.5).

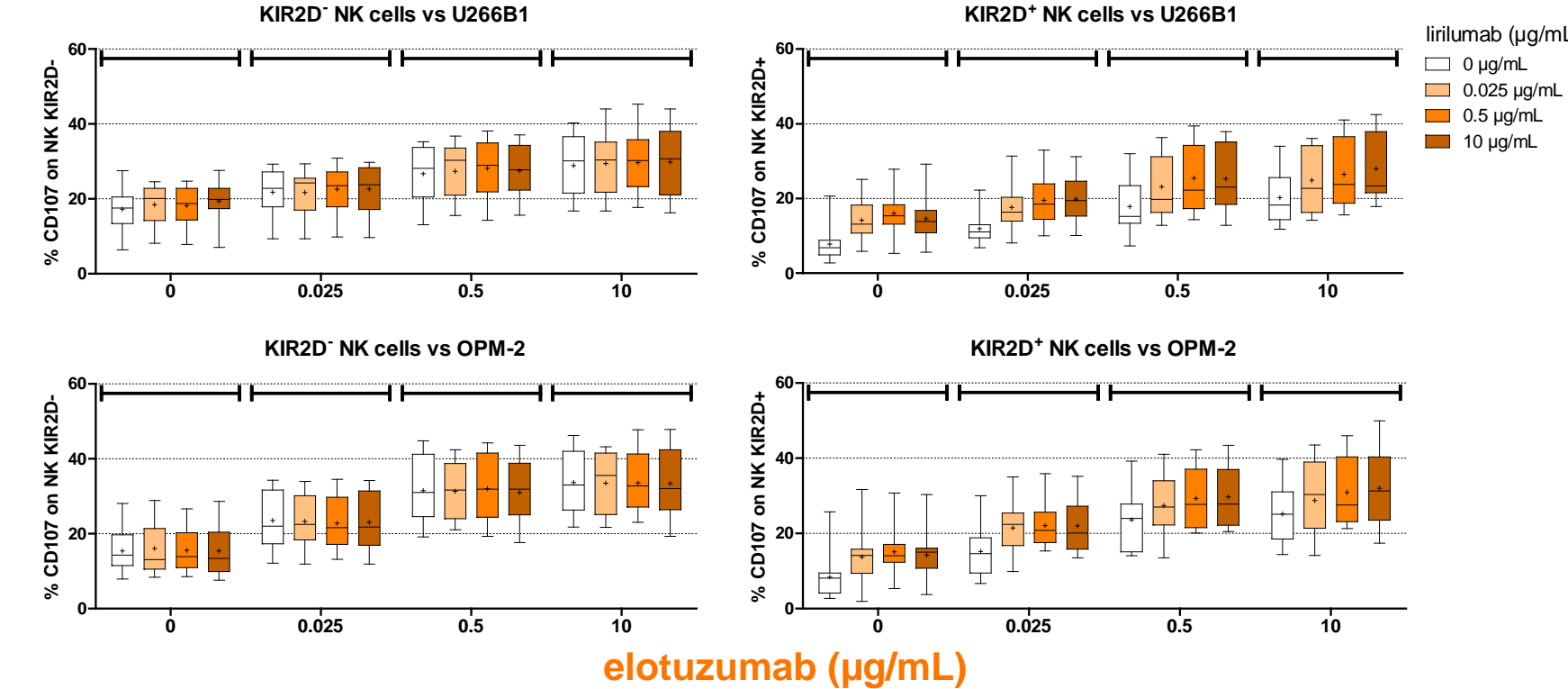
**Table 1: Characteristics of PBMC isolated from 12 healthy volunteers**

Donor ID	CD16 typing	HLA-C typing	% NK in PBMC	% lirilumab positive NK cells
1	F/V	C1/C2	18%	76%
2	F/F	C1/C1	7%	37%
3	F/F	C1/C2	7%	52%
4	F/V	C1/C2	10%	42%
5	F/V	C2/C2	10%	33%
6	F/F	C1/C1	19%	52%
7	F/V	C1/C2	28%	59%
8	F/F	C1/C2	14%	68%
9	V/V	ND	8%	38%
10	V/V	C1/C1	4%	39%
11	V/V	ND	10%	51%
12	V/V	C1/C2	22%	43%
		<b>Mean</b>	<b>13%</b>	<b>49%</b>
		<b>SD</b>	<b>7%</b>	<b>13%</b>
		<b>Median</b>	<b>10%</b>	<b>47%</b>
		<b>Range</b>	<b>4-28%</b>	<b>33-76%</b>

- *In vivo* assessment of the therapeutic efficacy of lirilumab and elotuzumab was performed in transgenic mice expressing human KIR2DL3 (results reported in poster 4717) and as reported in this poster in a novel strain of double-transgenic mice we developed, expressing human KIR2DL3 as well as its ligand, HLA-cw3, on a Rag1<sup>-/-</sup> background (KIR-cw3-tg RAG mice). The OPM-2 MM cell line was subcutaneously engrafted in these mice and when high tumor volumes were reached (>100mm<sup>3</sup>), mice were treated as indicated (10 mice per group).

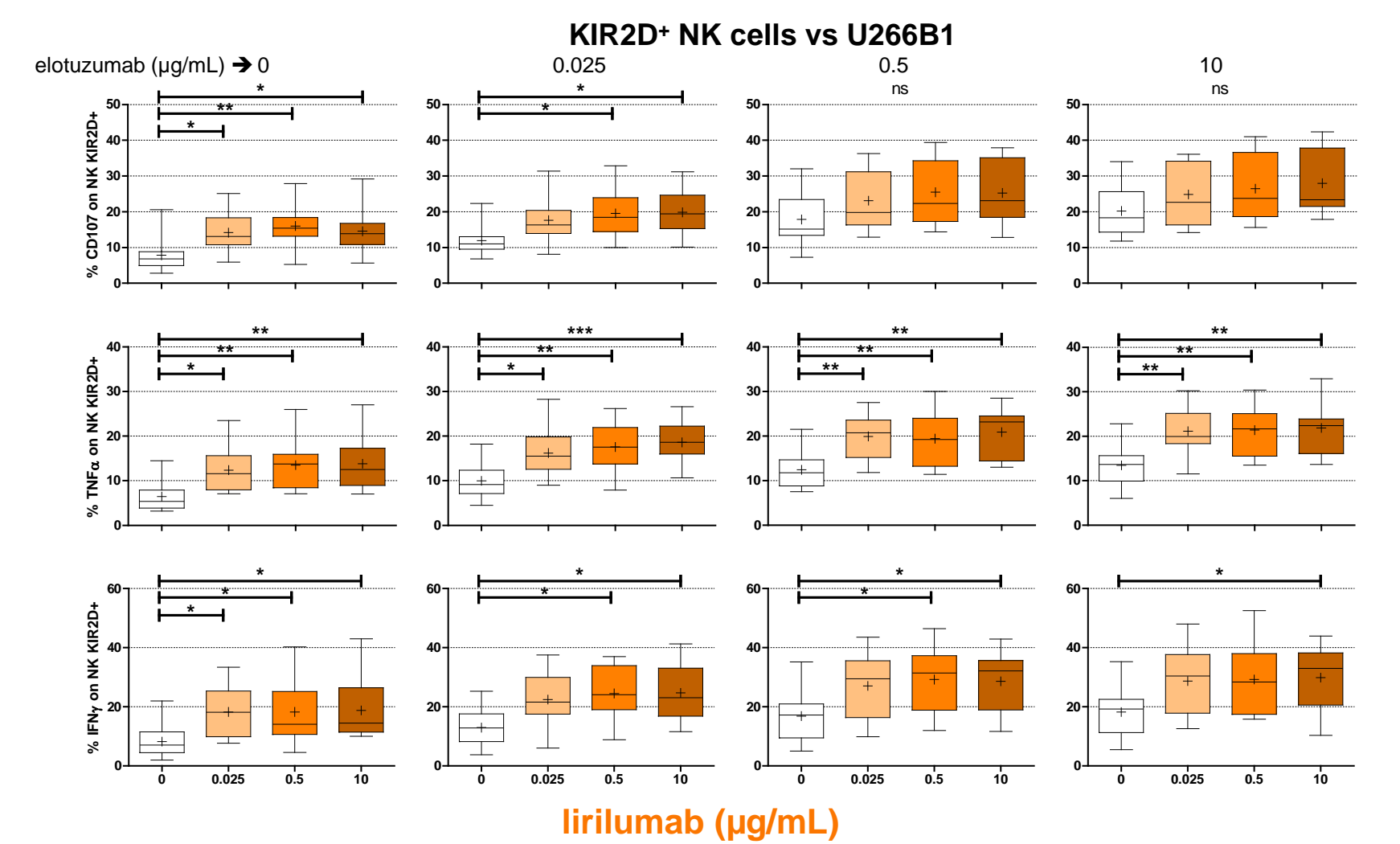
## Results

**Figure 2: Lirilumab restores specifically KIR2D<sup>+</sup> NK cell activation towards multiple myeloma cell lines U266B1 and OPM-2 *in vitro*, in a dose dependent manner, and enhances elotuzumab-mediated ADCC**



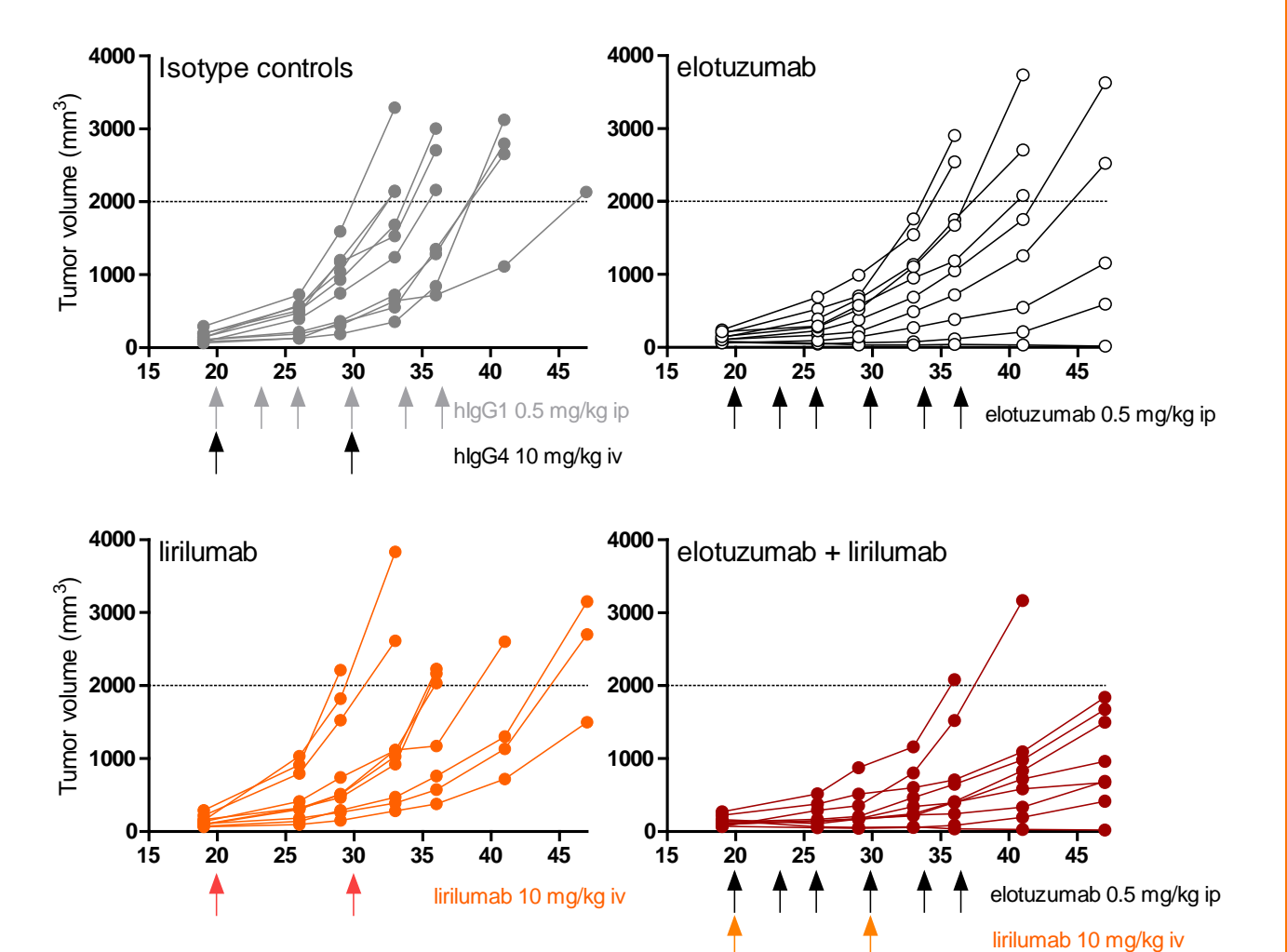
**Fig. 2:** CD107 mobilization on NK cells expressing KIR2D<sup>+</sup> (KIR2D<sup>+</sup> NK cells, right column) targeted by lirilumab or on NK cells not expressing KIR2D (KIR2D<sup>-</sup> NK cells, left column). CD107 mobilization is measured on indicated NK cells, within PBMC incubated with U266B1 cells (top panel) or with OPM-2 cells (bottom panel) for 4h at 37°C. Box plots indicate the median with the second and third quartiles. Whiskers indicate min. and max. values. + indicates the mean. N=12 healthy volunteers. Similar results were obtained by measuring intracellular cytokines (TNF-α, IFN-γ) contents (data not shown).

**Figure 3: Combined effect of lirilumab and elotuzumab on KIR2D<sup>+</sup> NK cells towards multiple myeloma cell line U266B1 is seen *in vitro* on CD107 mobilization, and intracellular cytokines (TNF-α and IFN-γ) production**



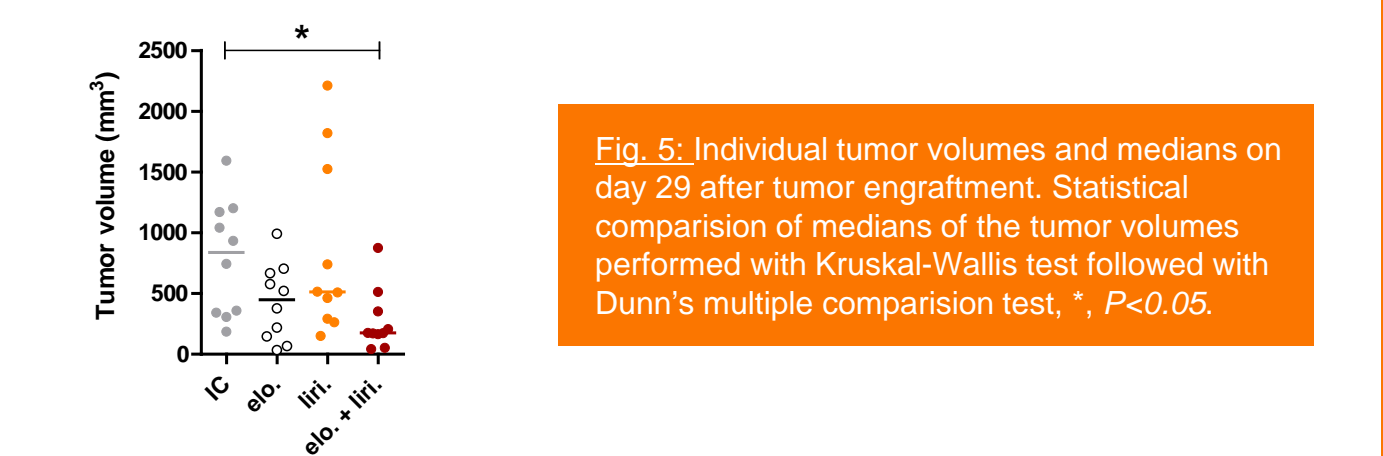
**Fig. 3:** CD107 mobilization (Top), intracellular TNF-α production (Middle) and intracellular IFN-γ production (bottom) in NK cells expressing KIR2D in response to U266B1 cells in 4h at 37°C, with indicated doses of lirilumab and elotuzumab. Box plots indicate the median with the second and third quartiles. Whiskers indicate min. and max. values. + indicates the mean. N=12 healthy volunteers. Groups were compared using a one-way ANOVA and a Bonferroni multiple comparison test. ns = non significant, \* p<0.05, \*\* p<0.01, \*\*\* p<0.001. Similar results were obtained with OPM-2 multiple myeloma cells. No significant effect of CD16 genotyping was observed in these experiments (data not shown).

**Figure 4: Lirilumab enhances *in vivo* anti-tumor efficacy of elotuzumab**



**Fig. 4:** KIR-cw3-Tg RAG mice were sc engrafted with 5x10<sup>6</sup> OPM-2 cells with matrigel. 19 days post tumor engraftment, mice were randomized for a mean tumor volume of 130 mm<sup>3</sup> (n=10 mice/group) and treated as indicated with arrows. Each curve corresponds to one individual tumor growth. Data are representative of two independent experiments.

**Figure 5: Combination therapy with lirilumab and elotuzumab significantly decreases tumor volumes**



**Fig. 5:** Individual tumor volumes and medians on day 29 after tumor engraftment. Statistical comparison of medians of the tumor volumes performed with Kruskal-Wallis test followed with Dunn's multiple comparison test, \*, P<0.05.

## References

Abstract #4717. Effects of IL-21, KIR Blockade, and CD137 Agonism on the Non-Clinical Activity of Elotuzumab.

## Conclusions

We demonstrate that blockade of KIR checkpoint receptors with lirilumab was able to augment elotuzumab mediated ADCC *in vitro* and synergized with elotuzumab to mediate potent anti-MM activity *in vivo*.

These data suggest that lirilumab treatment may increase the therapeutic efficacy of elotuzumab:

- *In vitro* MM cells were capable of activating peripheral blood NK cells from healthy donors, responses were significantly enhanced, in a dose-dependent manner, by both lirilumab and elotuzumab independently
- The elotuzumab-mediated functional activation of KIR2D<sup>+</sup> NK cells could be further enhanced by the addition of increasing doses of lirilumab.
- *In vivo*, as monotherapy, each of monoclonal antibody had some therapeutic effect while the combination of both resulted in a significantly stronger anti-tumor effect and increased survival of the mice.

Taken together, these data provide a rationale for clinical trials to test combination treatment of lirilumab and elotuzumab in MM patients.